



## Taking the Pulse of Natural History Collections During COVID-19 Series: Where are we now?

### September 16: Collections' perspectives

***John Bates, The Field Museum***

***Emily Braker, University of Colorado Boulder***

***Brian Atkinson, The University of Kansas***

***Mare Nazaire, California Botanic Garden***



## The Field Museum in the time of Covid-19





# The New Museum “Normal”

- Decreased attendance
  - Budget shortfalls
  - Staff reductions
  - Salary reductions
  - Frozen searches
- 
- Little collections access
  - Reduced lab access
  - No fieldwork
  - Working from home
  - Zoom fatigue
  - Reopening strategies

# The Field Museum: 126 years of Biodiversity collections and research









Expertise

# The “New” Museum

- Working from home (the upside)
- Honest discussions of I.D.E.A.
- Virtual Meetings (global access)
- Virtual Educational Activities  
(broader local and regional access)
- Virtual interactions with donors (no need to drive to the museum)
- Realizing the benefits of digitization



# Issues of societal concern.



Closed for the pandemic, the Field Museum of Natural History hosted a socially-distanced blood drive in its empty, cavernous halls. SCOTT OLSON/GETTY IMAGES

## Shuttered natural history museums fight for survival amid COVID-19 'heartbreak'

By Elizabeth Previdi | May 28, 2020, 8:55 PM



# Inclusion, Diversity, Equity, and Accessibility: So much to do



# ¿Cómo pueden surgir especies por cambios en el comportamiento migratorio?

Valentina Gómez-Bahamón

**FIE**  
**LD.**

**UIC**

**SELVA**

John Bates

Carolina Casas

Maria Castaño

Valentina Gómez

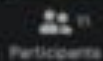
David Castaño



Unmute



Stop Video



Participants



Chat



Share Screen



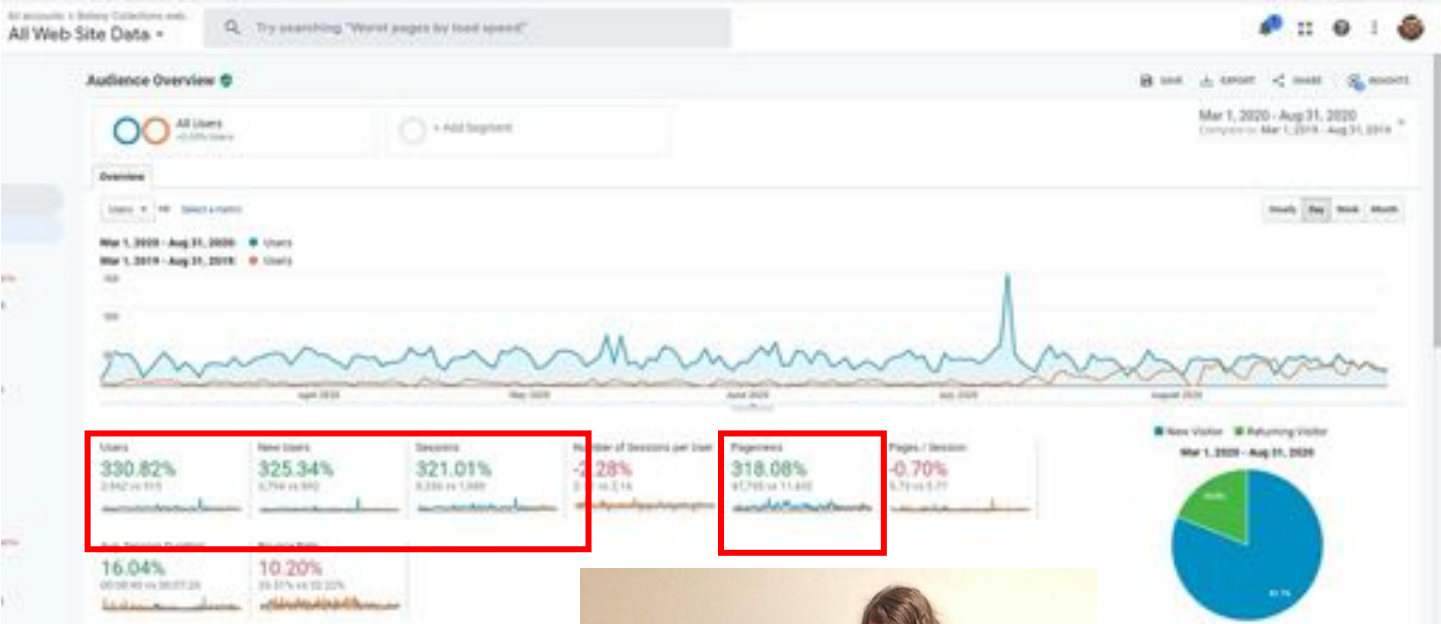
Record



Reactions

Leave

# Field Museum Botany: Increased access of the web site





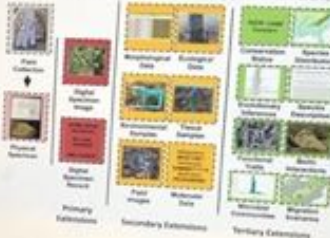
## The Extended Specimen

### The Extended Specimen

Emerging Frontiers in Collections-Based Ornithological Research



### Extended Specimens in Action: Appalachian Lichens



Lendemer et al. 2020

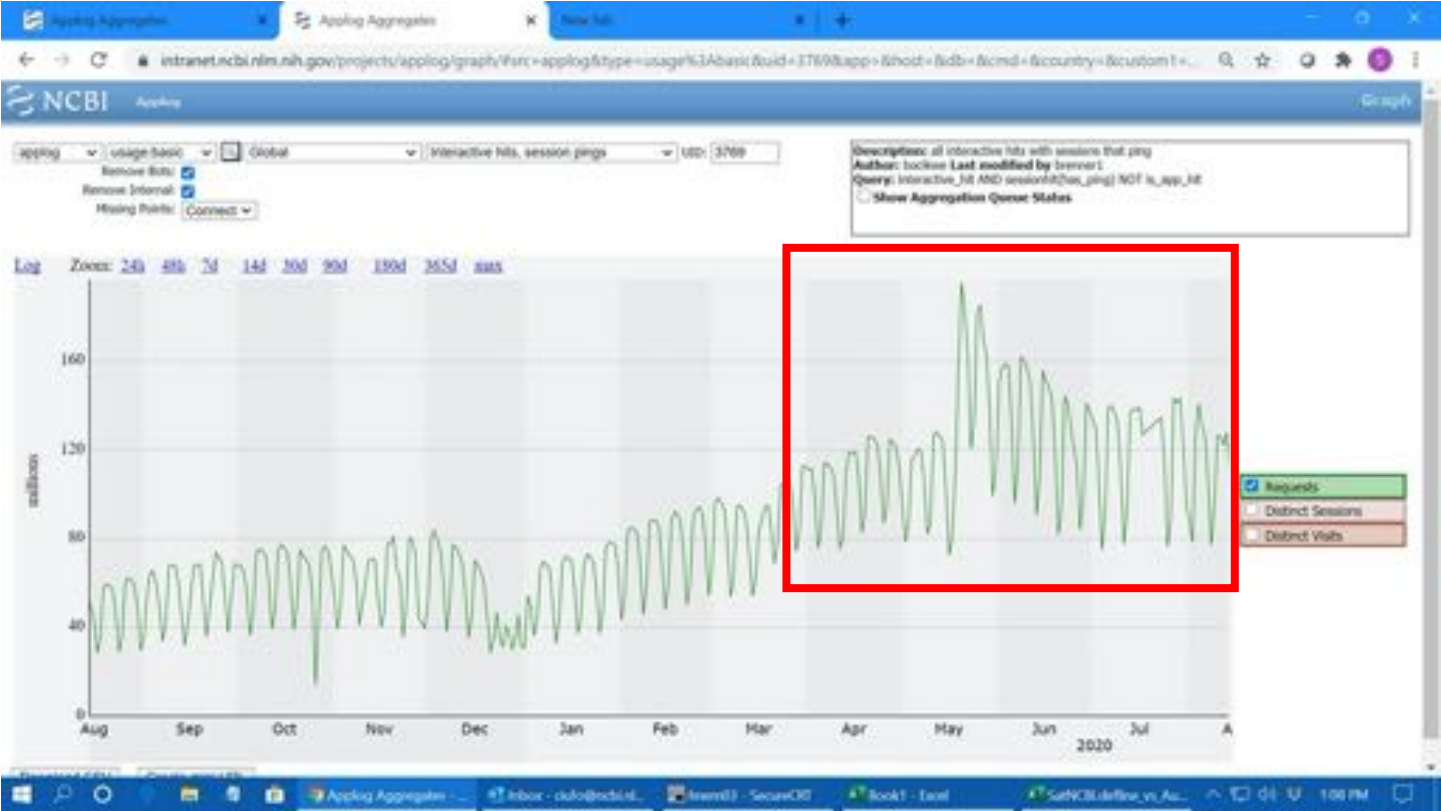


## The Extended specimen: Genotype

*Long-term archival of genetic resources available at multiple facilities (thousands/millions of specimens) for scientific research*



# GenBank sequence downloads for the last 12 months





# The extended specimen realized: phenotype

NSF supported CT-scanning of vertebrates

oVert TCN Project Description

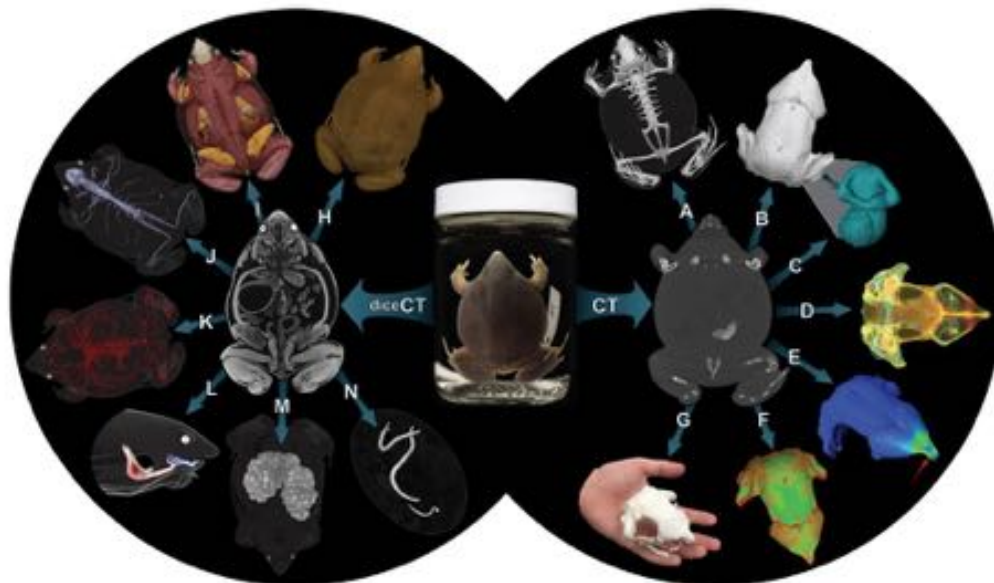
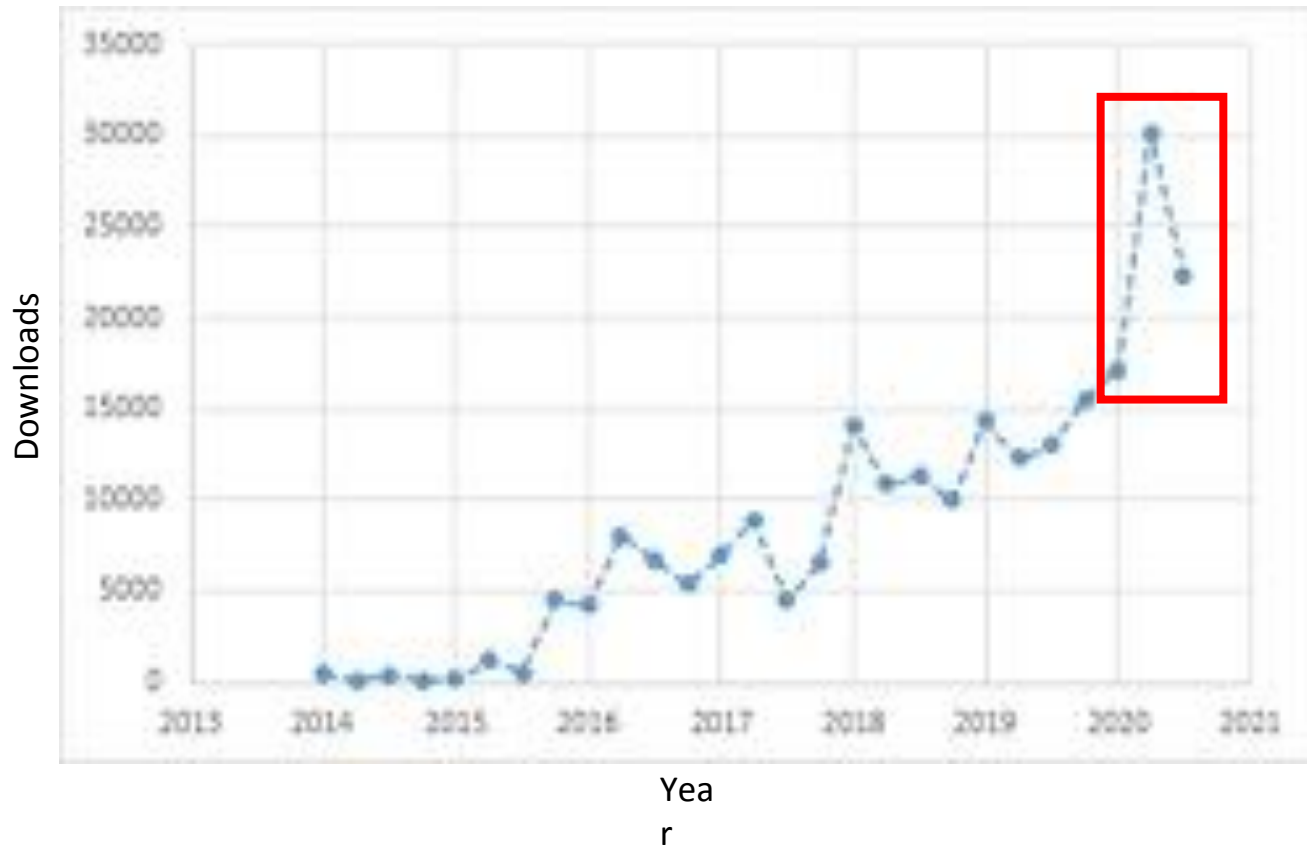


Figure 1. Diversity of data and analyses resulting from traditional computed tomography (CT) and diffusible iodine-based contrast-enhanced CT (diceCT) of a formalin-fixed, ethanol-stored frog specimen (genus *Hemisus*). A, qualitative morphology of skeleton; B, isolated cranium with, C, inner ear endocast; D, density analysis; E, Finite Element Analysis (10) showing distribution of stresses when force applied (red arrow); F, wall thickness analysis; G, 3D printed model of skull; H, external surface anatomy (similar to photogrammetry of specimens); soft-tissue anatomy including, I, skeletal muscles and glands, J, nervous system, K, cardiovascular system, and, L, intrinsic tongue muscles (blue, m. genioglossus; red, m. hyoglossus); natural history 'by-catch': M, eggs in oviducts, and, N, endoparasites (nematodes).



# Quarterly downloads of Morphosource 3-D specimen data



# Our “New” Museum

- Working from home if you can
- Virtual Meetings (global access)
- Virtual Educational Activities  
(broader local and regional access)
- Virtual interactions with donors  
(no need to drive to the museum)
- Continued digitization
- Beyond discussions of I.D.E.A.





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Museum of Natural History

UNIVERSITY OF COLORADO BOULDER

# COVID Operations and Interim Outcomes

## VERTEBRATE COLLECTIONS

University of Colorado Museum of Natural History

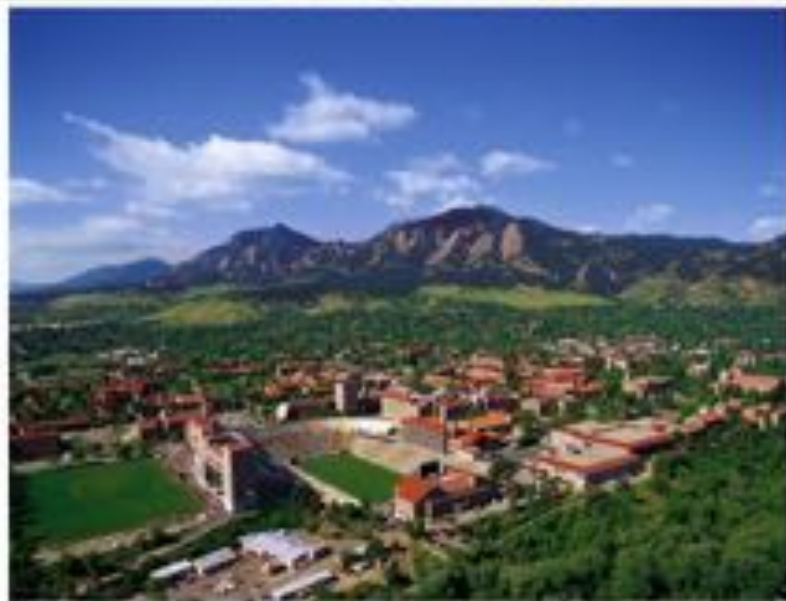




# Museum of Natural History

UNIVERSITY OF COLORADO BOULDER

- Boulder, Colorado
- Student population ~35,000
- 5 million natural history specimens
  - Anthropology
  - Botany
  - Diatoms
  - Entomology
  - Paleontology
  - Zoology
- 50,000 visitors annually. Audiences:
  - K-12
  - University students
  - General public
- Museum and Field Studies (MFS) Graduate Program





## Current campus operating mode = **EXPANDED**

- Limited in-person research and teaching
- Online course offerings
- Remote work for noncritical staff
- Infrastructure mitigation - HVAC improvements
- Formal 'return to campus' request process
- Daily online health assessment
- Limited COVID testing (dorms, waste streams)



Stay 6 feet apart.



*Mandatory mask wearing, social distancing, enhanced custodial and sanitation practices*



**HAVE YOU  
COMPLETED  
YOUR DAILY  
HEALTH FORM?**



[uclinks.edu/daily-health-form](https://uclinks.edu/daily-health-form)







# Preparation for Return to Campus

- Campus COVID Task Force coordination
- PPE and cleaning supply distribution
- Directional & safety signage
- Building security – after-hours campus police sweeps due to extended class schedule (7am-10pm)



# Vertebrate Collections – Current Operations

- Hybrid mode: In-house and remote tasks
- Hiatus on visiting researchers and volunteers
- Reduced occupancy: 1 person per room
- Increased sanitation practices





Positive  
Outcomes

# I. Priority Reshuffling: *Rise of the backburners*



- Collections Policy revision
- Data improvement projects
  - Georeferencing
  - Taxonomy updates
  - Data Linkages:
    - Citations
    - GenBank sequences
    - Exchanged materials
  - Media Linkages:
    - Ledger pages
    - Field notes



**UCM:Mamm:8166** On Smith Trail, near Coos Creek, 2 miles W of Cresto  
North America: United States, Colorado, Gunnison County  
11 July 1962 (1962-07-11)

**Arctos**

**IDENTIFICATION** | **COLLECTOR** | **IDENTIFIERS** | **LINKS**

**IDENTIFICATION** | expand | **Marmota flaviventris futeola**  
Specimens: Charles S. Townsend, Frederick S. Townsend, Bernard, Marmota flaviventris futeola  
Identified by Edward R. Warren on 1962  
Nature of ID: Features  
Confidence: High  
Remarks: Former nature\_of\_id type specimen  
Locality: futeola.html

**COLLECTOR**  
collector: Edward R. Warren

**IDENTIFIERS**  
ARK: 63665/16689402-5967-4657-4021-4c56c256a1ed  
collector number: ERM 35  
original identifier: 106204  
USNM: National Museum of Natural History, Smithsonian Institution  
202937

**CITATIONS** | **PERSISTENT IDs, URLs**

Citations  
viewer of Marmota flaviventris futeola in Warren 1962 (10-11-2017) (1962) |  
Phototype of Marmota flaviventris futeola - page 18 in Howell 1914

Links  
GBIF Occurrence

## II. Increased Collaboration

- Collections Managers



- Sharing strategies
- Continuity of Operations Planning (COOP)
- Collaborative student training

- Institutional

- Virtual Exhibits
- MFS virtual tours and panel discussion
- Museum Unlocked podcast
- Gallery space for in-person classes



*Museum gallery Mammalogy Lab – gloves required, traffic guides on floor, socially distanced lab stations*

### III. Opportunistic Continuity Tools

- Video Tutorials
  - Zoom recordings
  - Annotated specimen prep videos
- COOP – Hotspot Maps
- Remote Work – possible, even for collections people!



Hotspot Map



## Lessons & Tips

- Consider collaborative training opportunities
- Video conferencing
  - Regular check-ins with remote staff
  - Record - Good for current and future technicians
  - Even when in person, database training is easier over Zoom (for social distancing)
- Scan archives! University collections may be able to partner with libraries
- Invest in laptops for staff when possible
- **Adaptability**



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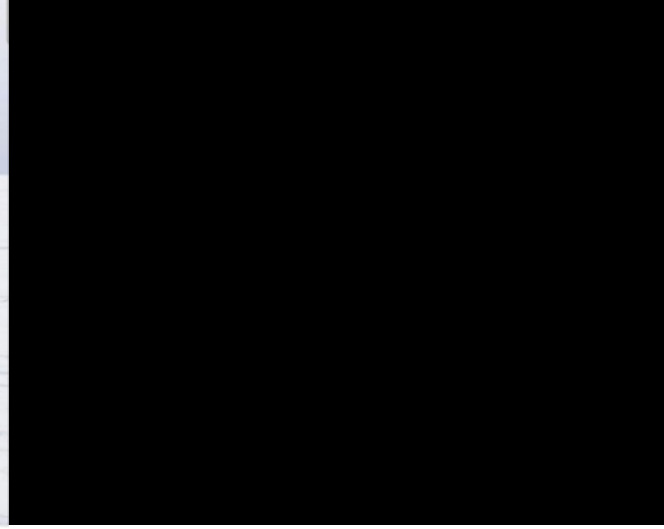
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# COVID-related operations from the perspective of a faculty curator teaching classes with a virtual lab

**Brian Atkinson**

Asst. Professor, Ecology and Evolutionary  
Biology Department

Curator, Division of Paleobotany, Biodiversity  
Institute

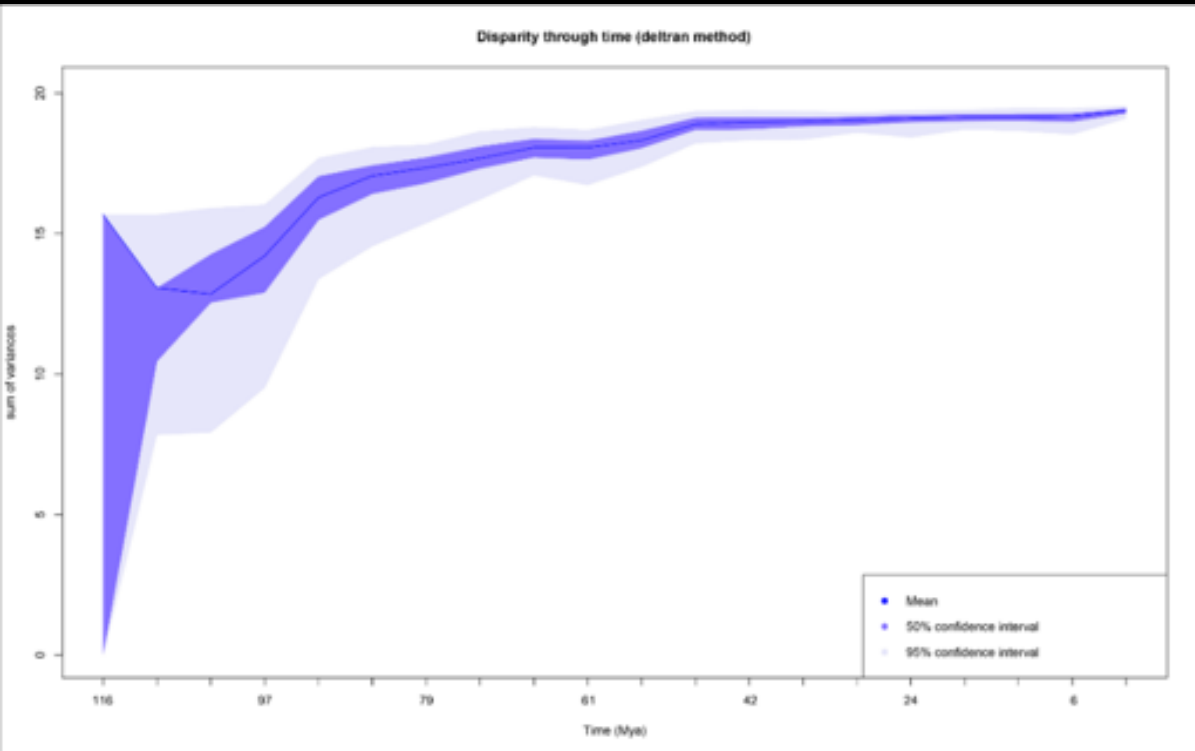
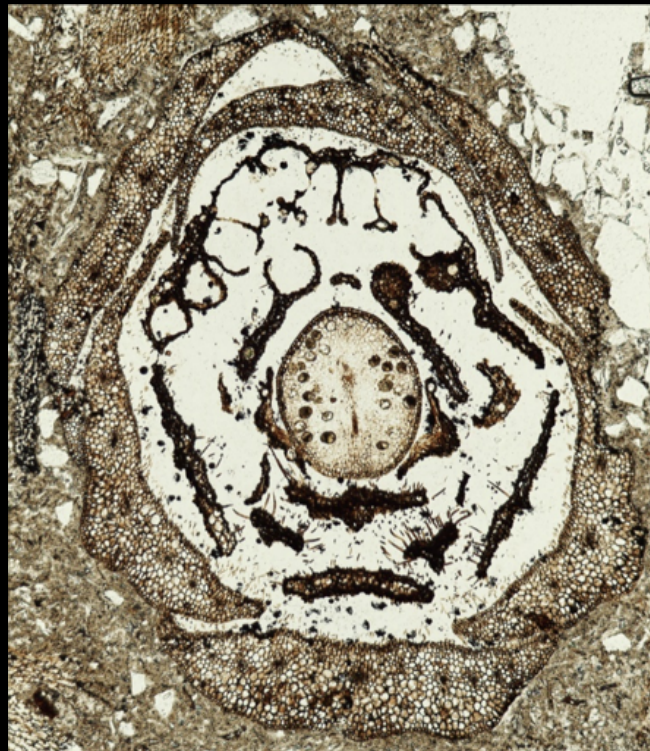
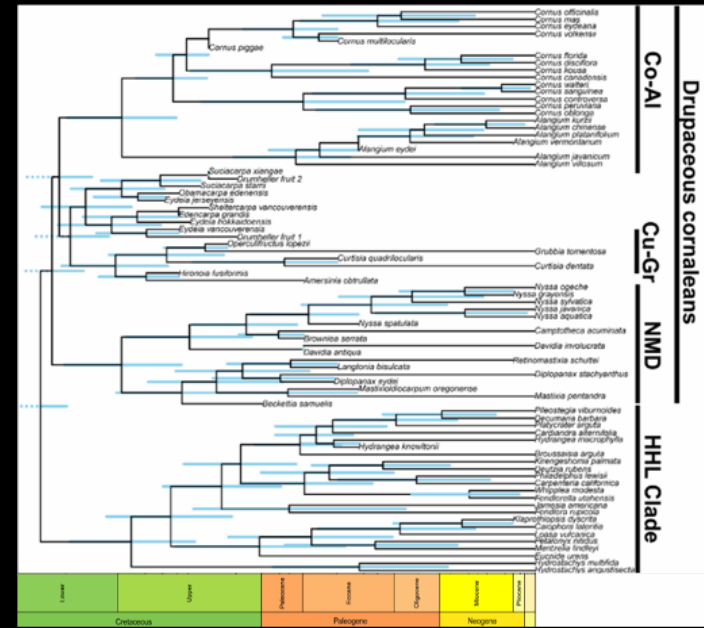
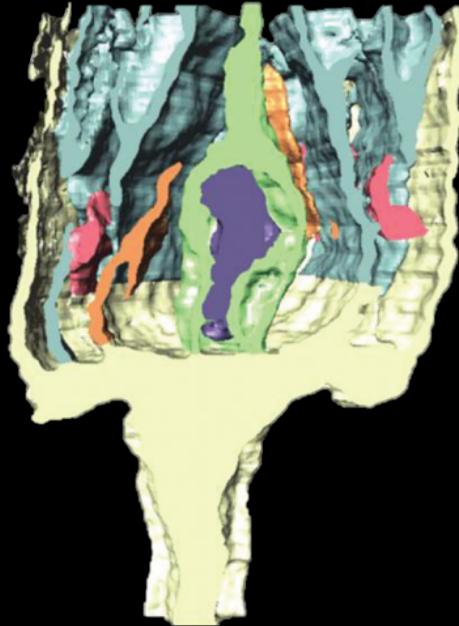
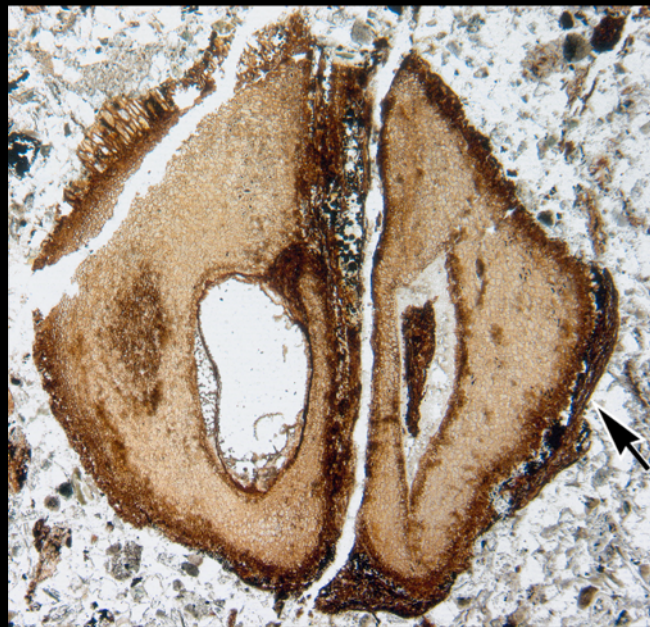
University of Kansas

Email: [brian.atkinson@ku.edu](mailto:brian.atkinson@ku.edu)

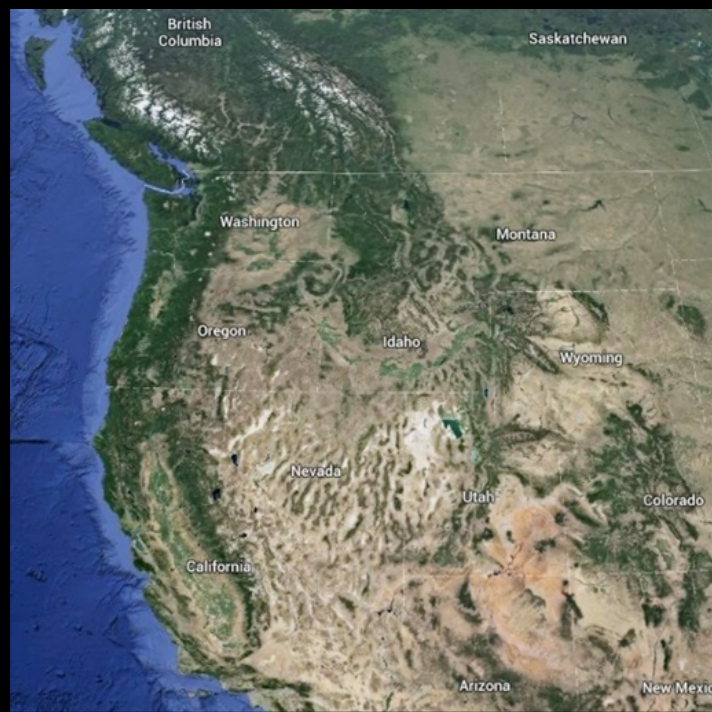


## **Break down of this report:**

- a. Introductions: myself, KU Paleobotany personnel, research, and collections**
- b. Status of KU Paleobotany before and during covid**
- c. Teaching a lab based course in a virtual format**
- d. Outlook**



# Active field program along the western coast of North America and Antarctica





Kelly Matsunaga  
Asst. Professor &  
Curator



Kelly Pfeiller  
PhD student



Ben Perry  
Undergraduate



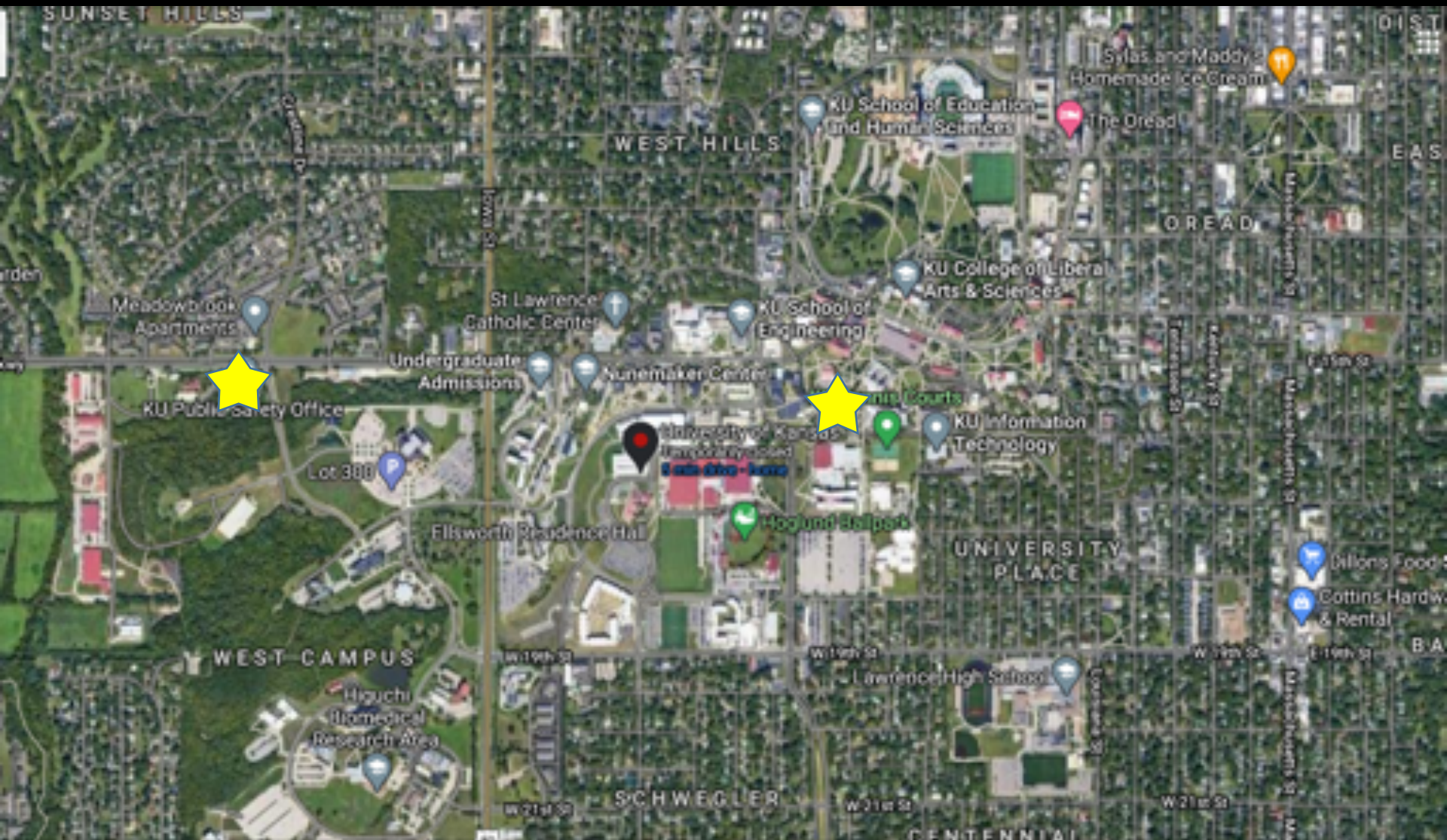
Anna Krause  
Undergraduate



Keana Tang  
Masters student



Rudolph Serbet  
Collections Mngr



## Collection and Research Strengths:

400,000 specimens: Pre-Cambrian-Recent

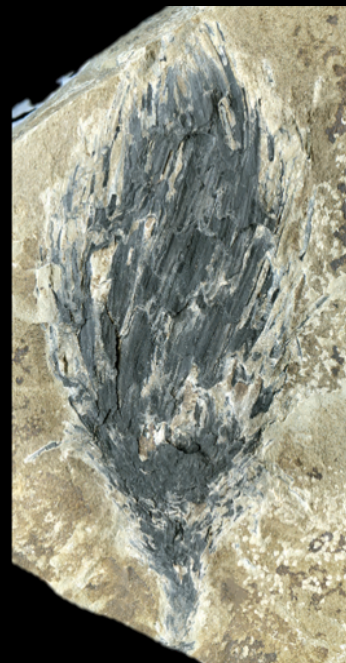
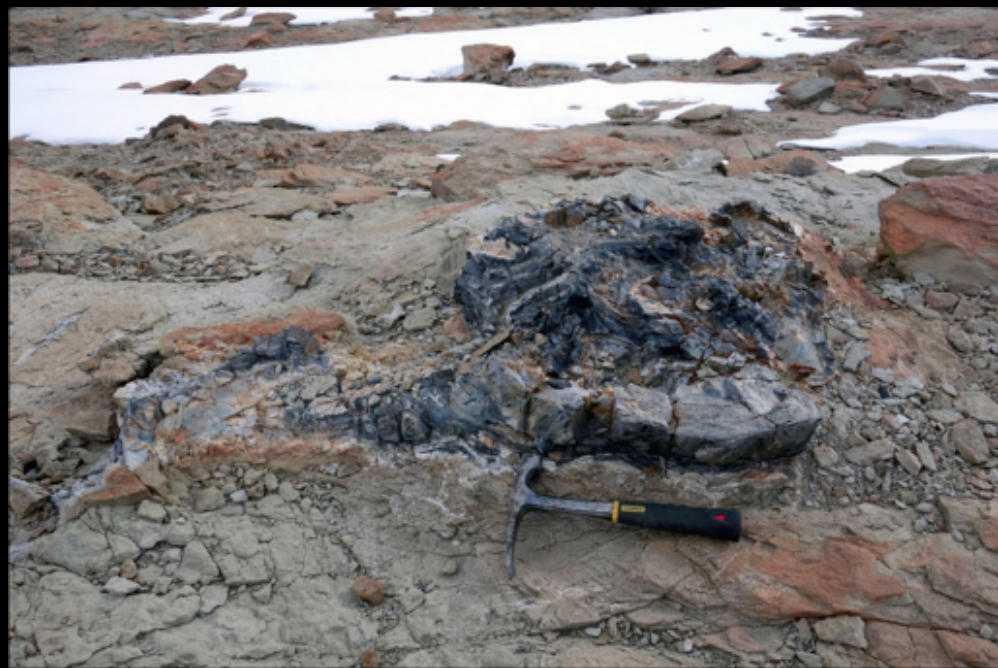
Largest North American repository for Antarctic fossil plants (>35,000 specimens, Permian-Jurassic)

NSF Sanctioned Repository for Antarctic fossil plants

Anatomically preserved plants from crucial time intervals

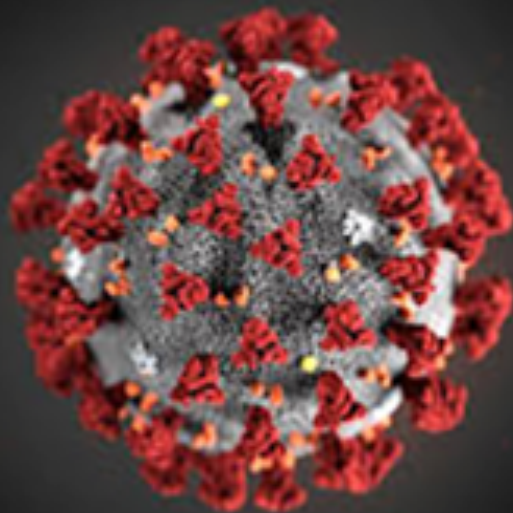
Currently digitizing our specimens via Specify











## **COVID-19, the Novel Corona Virus**

**News and information for the ERA-EDTA  
community and kidney patients.**

Managing a research program and collection in the  
middle of a pandemic???

## Research within the division:

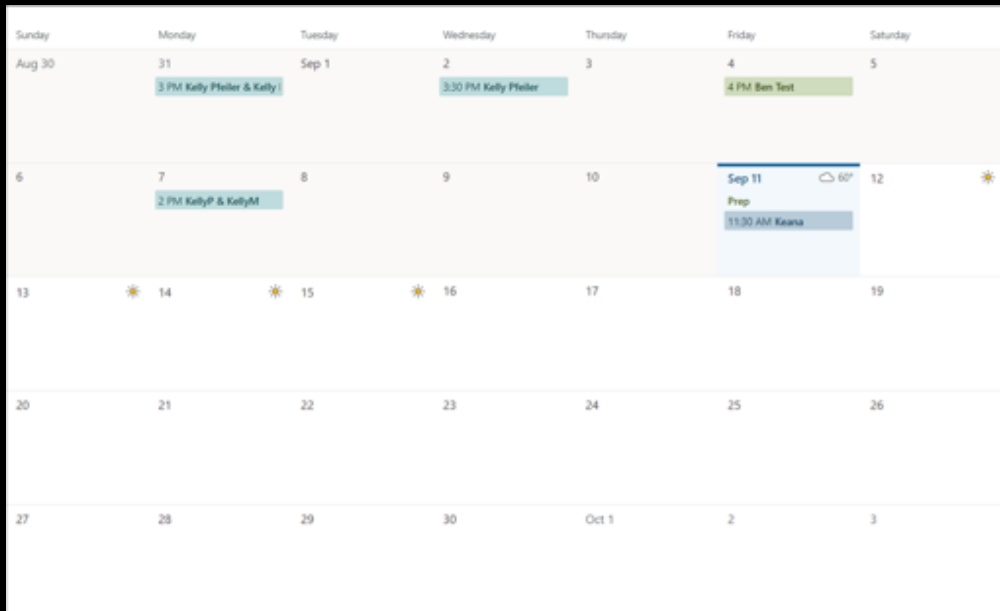
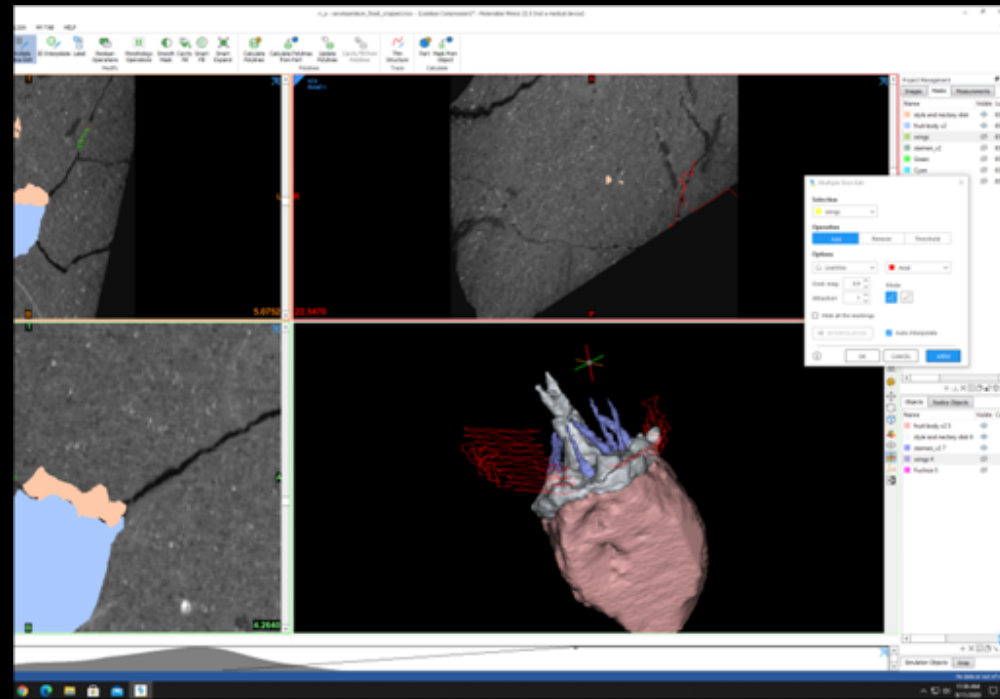
Everyone has scopes at home

Remote access to fast computers for Ct-imaging processing

Dedensifying the lab

## Collections management:

Loans are still out and normal operations have resumed



# Teaching

Diversity and History of Organisms (lecture and lab)

Co-taught: Myself (everything except Metazoa); Rich Glor (Metazoa)

Lab has traditionally been a hands-on specimen based experience



Ray Troll



Teaching a biodiversity class in the middle of a pandemic???



# Phylogenetic Tree Literacy

Given the current situation  
and a prioritization of  
safety. What do we want  
the students to walk away  
with this year?



Richard Glor  
Professor &  
Curator



Walter Taponjou  
PhD Student &  
Graduate TA



Keana Tang  
PhD Student &  
Graduate TA



Anna Krause  
Undergrad Rschr.  
& Undergrad TA

# Asynchronous pre-recorded lectures

## Synchronous recitations, 2 sections per week

### Open book exams (24 hrs to take exam)

The screenshot shows the Canvas LMS interface for the course '2020Fall-BIOL 413 History and Diversity of Organisms LEC 4209-10395'. The page title is 'Weekly Modules'. On the left, there is a navigation menu with options like 'Announcements', 'Resources for building your course', 'Syllabus', 'My Grades', 'Pre-Course', 'Week 1', 'Weekly Modules', 'Phylogenetic Tree', 'Reading Resources', 'Exams', and 'Additional Study Materials'. The main content area shows a list of weekly modules:

- Week 1**
- Week 2** Enabled: Adaptive Release, Statistics Tracking
- Week 3** Enabled: Statistics Tracking
- Week 4** Enabled: Statistics Tracking
- Week 5** Enabled: Statistics Tracking

At the top of the main content area, there are tabs for 'Build Content', 'Assessments', 'Tools', and 'Partner Content'.

The screenshot shows a video playlist interface with several lecture thumbnails:

- Week 4** Updated September 7th, 2020. Playlist Details. Thumbnail: Eukaryotes (Phylogenetic Tree)
- Week 3** Updated August 31st, 2020. Playlist Details. Thumbnail: DIVERSITY AND LIFE CYCLE (Microscopic images)
- Week 1** Thumbnail: Billion Years Ago (Timeline)
- Week 1** Thumbnail: INTRODUCTION TO CELLULAR LIFE (Microscopic images)
- Week 1** Thumbnail: VIRUSES (Microscopic images)

Each thumbnail includes a title, a brief description, and a small video player icon.



# Online labs:

## Tree thinking exercises using iTOL

## Hopefully: diversity surveys via iNaturalist

### Lab 1: Viruses & the Three Domains

**Introduction**

This week's lab explores phylogenetic trees for viruses and the three major domains of non-viral life. In Part 1, you will do some basic tree reading exercises to investigate the origins of our current pandemic. One important goal of these exercises is to help you better understand monophyly, paraphyly and polyphyly, and their evolutionary implications. By the end of these exercises you should be able to interpret important phylogenetic data about the origins of COVID-19 better than some authors who have published on this topic. In Part 2, you will use a series of exercises to explore a tree for the three major domains of non-viral life (Archaea, Eukaryota and Bacteria). You will use this tree to gain experience with important basic tree concepts like most recent common ancestor (MRCAs), tree rooting, and tree support. Finally, you will learn how to map traits onto your tree and identify different types of homologous traits (e.g., synapomorphies, autapomorphies and symplesiomorphies).

**Goals**

**Exploring the tree of life**  
Use phylogenetic trees to trace the origin of covid-19 and examine relationships between the three domains of life (Archaea, Eukaryota and Bacteria)

**Concepts**  
Visualization and interpretation of different types of phylogenetic trees; monophyly, paraphyly, and polyphyly; mrcas, homology, autapomorphy, synapomorphy and symplesiomorphy

### Part 1: COVID cobras and pangolins: using phylogenetic trees to trace the zoonotic origins for SARS-CoV-2

**Background:** The Spanish flu of 1918, ebola, SARS, and now COVID-19, all result from viruses that jumped to humans from another animal species. These infectious agents are known as **zoonoses** (i.e. zoonosis). Determining the sources zoonoses is critical to understanding how pathogens evolve, and potentially stopping zoonotic transfers from occurring in the first place. Phylogenetic trees are one of the main tools that scientists use to identify the sources for zoonotic viruses. The simplest approach is to build a phylogenetic tree that includes the human virus together with related viruses from other species of animals; this allows scientists to identify the likely source species as the one hosting the virus that is most closely related to human pathogen. In this manner, the Spanish flu of 1918 has been traced to birds (Hoog 2014) and Middle Eastern Respiratory Syndrome (MERS) to domesticated dromedary camels (Wang et al. 2020).

Two recent global outbreaks of Severe Acute Respiratory Syndrome caused by coronaviruses – first SARS and now COVID-19 – appear to have emerged initially at live animal markets in Asia, where a remarkably diverse range of animal species are regularly bought and sold. SARS, which killed

Fig. 1. Headlines from January 2020 suggesting that SARS-CoV-2 jumped to humans from snakes.

**Unit 14.6**

of "bacteria" was a major discovery with important implications for the evolution of life on earth, but reconstructing the early evolution of life's three domains has always been complicated by the absence of traits that can be used to distinguish between the two different types of "bacteria," or to determine which of these groups is most closely related to eukaryotes. In this exercise, we will examine relationships across the three domains of life, as well as the distribution of traits that distinguish and define these groups. Along the way, we will learn about important differences between rooted and unrooted phylogenetic trees, how to identify the traits that distinguish and define major groups of life, and other basic lessons about reading and interpreting phylogenetic trees.

**Part 2 Goals:** Open a phylogenetic tree using online software. Identify most recent common ancestor (MRCAs) and shared branches. Learn the important differences between rooted and unrooted trees; interpret tree support values. Classify traits as homologous, synapomorphies, symplesiomorphies, and autapomorphies.

**Exercise 3: Identify MRCAs and shared branches for major domains of life**

**Background:** You must master the ability to recognize MRCAs in this course. This exercise will provide a lot more practice with this important task.

**Exercise 3 Goals:** Identify MRCAs on a tree illustrating relationships between the three domains of life.

1. Open a tree illustrating the three domains of life in iTOL. Go to <https://itol.med.utah.edu/>. Click the Tree of Life tab at the top of the page. You should see a circular tree with colored lines around its tips (Fig. 4). These colored lines indicate the three main domains of life: Archaea, Bacteria, and Eukaryota. You can see which colors correspond with which clades either by finding and expanding the Colored ranges pop-up window (Fig. 4C), or by moving the mouse over the tips of the tree, which should result in pop-ups that give species names, links to additional information, and, in some cases, a photograph (Fig. 5).
  2. Use the Manual annotations option in iTOL (Fig. 6A) to draw a circle around the most recent common ancestor (MRCAs) of Eukaryota and Archaea.
  3. Use the Draw a rectangle/square tool in the Manual annotations tools pop-up window to put a square in the middle of the branch that separates Eukaryota + Archaea from Bacteria. In this case, you can hold **Alt** when creating your symbol to get a square rather than a rectangle.
  4. Use iTOL to generate an image to submit with your lab report. Go to the Controls (Fig. 4E) pop-up window and select the Export tab. Select PDF. Portable Document Format from the Format dialogue box. Click Export.
- Blackboard:** Submit PDF image of tree containing annotations.

**Fig. 8. The unrooted tree of life in iTOL.**

**Exercise 4: Tree Rooting**

**Background:** All of the trees that we have viewed during this lab so far have included an assumed node known as the root, which represents the MRCA for all of the taxa in the tree. Although identifying the root of a phylogenetic tree is critical to answering important evolutionary questions, most tree-building algorithms generate unrooted trees that require the addition of a root using additional information and methods. The most frequently used method for rooting phylogenetic trees involves designating one or more outgroups, or taxa that are known a priori to fall outside of a particular focal clade (ingroup). For example, if we are interested in inferring the root of mammals, we might also include a few other vertebrates as outgroups, inferring the location of the root of our focal clade is complicated because we do not know which of the three domains is the outgroup. In the trees we have been looking at in iTOL, the root separates Bacteria from a Archaea+Eukaryota clade. Our goal here is to explore the differences between rooted and unrooted versions of our tree for the three domains of life.

**Exercise 4 Goals:** Learn about the important differences between rooted and unrooted phylogenetic trees.

1. Go to the Controls pop-up in iTOL (Fig. 4E), click the Basic tab, and change the Display mode from Circular to the rooted (Fig. 7).
  2. Use the Draw an ellipse/oval tool to put a circle around the most MRCA of Eukaryota and Archaea.
  3. Use the Draw an ellipse/oval tool to put a rectangle around the MRCA of Eukaryota and Bacteria.
- Blackboard:** Eukaryotes share a MRCA with Archaea than with Bacteria in this unrooted tree.
4. Use the Add a text label tool under the Manual annotations box to write the word "root" on the branch where the root would occur if Bacteria were most closely related to Eukarya than to Archaea.
  5. Use iTOL to generate an image to submit with your lab report. Go to the Controls (Fig. 4E) pop-up window and select the Export tab. Select PDF. Portable Document Format from the Format dialogue box. Click Export.
- Blackboard:** Submit PDF image of tree containing annotations.

**Unit 14.7**

method for assessing how well-supported individual nodes in the tree are. We are not going to discuss these methods in detail here because this requires a detailed understanding of the underlying phylogenetic algorithms; for the purposes of this lab, all you need to know is that support values on individual nodes often range from 0 to 100 (or 0 to 1), with higher values indicating greater support. Interpretation of these support values is also nuanced, but as a general rule of thumb, nodes that are supported by values over 90 are considered well-supported while those supported by values less than 50 are considered poorly supported.

**Exercise 5 Goals:** Learn how to view and interpret support values on the nodes of phylogenetic trees.

1. Go to the Controls pop-up window in iTOL and select the Advanced tab.
2. Change the Bootstraps/Posteriori option from Hide to Display.
3. Look at the raw bootstrap values on your tree by selecting the Text option (the default is Symbol). Zoom in and investigate the support values across this tree.
4. Further investigate node support by clicking back from Text to Symbol and changing the display range from 90-100 (Fig. 8). This should show you a symbol only on the well-supported nodes. Now change the display range from 0.0 to 1.0 to see nodes that are poorly supported.

**Exercise 5: Node Support**

**Background:** Before we get any further into reading and interpreting trees, we need to learn about some important tools that will help prevent us from wasting time or making mistakes interpreting patterns that are not well supported by our data. Most phylogenetic tree building algorithms provide some way to indicate how well supported individual nodes in the tree are.

**Fig. 9. The iTOL tree of life with symbols indicating bootstrap support.**

**Fig. 10. The tree of life in Normal view with traits of tips. Trait key (open symbol/filled symbol):** 1. eukarya (absent/present), 2. archaea (absent/present), 3. cytochrome oxidase (absent/present), 4. chloroplast/cyanelle (absent/circular/semi-circular), 5. flagellum (simple/complex), 6. promoter (Phobos box/TATA box), 7. start codon (Formyl-methionine/methionine), 8. cell walls (peptidoglycan/no peptidoglycan), and 9. membrane lipids (unbranched hydrocarbon/branched hydrocarbon). A. This pop-up window appears when you add traits; you can save the changes if you create an account at iTOL.

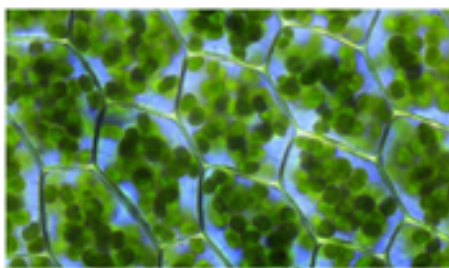
2 Reconstructing the history of traits that can be gained and lost using parsimony is not difficult, but does require us to learn a simple algorithm known as **Fitch parsimony**. Fitch parsimony was invented as a solution to solving parsimony questions a half century ago and remains widely used today. In this lab you will use Fitch parsimony's signature downpass/uppass algorithm to infer the evolutionary history of a game-changing evolutionary innovation: photosynthesis (Fig. 2). For details on using Fitch parsimony, see Box 1 below.

**Goals:** Learn how to use Fitch parsimony to reconstruct trait evolution.

## Exercise 1: Good idea! I'll take it!

**Background:** One important goal of this week's lab is to learn how to reconstruct the evolution of traits that may exhibit reversible evolution via Fitch parsimony. A secondary goal is to learn more about another explanation for why a trait might be scattered across the tree of life. Last week, we learned that complex multicellularity evolved repeatedly in distantly related groups via convergent evolution, and always from ancestors that had themselves independently evolved simple multicellularity.

A second way that organisms in different parts of the tree can obtain similar traits is via horizontal transfer, or, put another way STEALING (Fig. 3)! We will define horizontal transfer broadly as any instance where one organism ac-



**Fig 2.** "The world is green," is a well-known train among ecologists, and can easily capture the success of photosynthetic organisms. It is also a good drop line if you hear anyone disrespecting plants.



**Figure 2.** A sample of bryophyte diversity. The top two images show the liverwort, *Marchantia*. At the top right, we can see the elongated archaegonia (female sex organs) that contain the egg and eventually the sporophyte. The bottom image shows an expansive moss gametophyte bearing small unbranched sporophytes.

carried by the wind (rather than water as seen in their aquatic and marine algal relatives). In some cases, such as in the liverworts (*Marchantia*) the gametophyte grows specialized structures that raise the female sex organs (archaegonia) and their short-lived sporophytes.

Before going any further, we need to mention that land plants are divided into two main groups: the **bryophytes** and the **polysporangiophytes** (vascular plants) are the only survivors of this later group). **Bryophytes** are characterized by having a sporic life cycle with alternation of generations where the **gametophyte** is the dominant multicellular form (Fig. 1); in this group, the **sporophyte** is simple, short lived, unbranched, and has only one sporangium. **Polysporangiophytes** (polysporangio- "multiple sporangia" -phyte "plant"), meanwhile, are characterized by having a sporic life cycle with alternation of generations where the **sporophyte** is the dominant multicellular form. In this case the **sporophyte** is long lived, complex, branched, and has multiple sporangia (see Fig. 2). As **polysporangiophytes** began to become more abundant and diverse, competition and natural selection drove increases in height that allowed plants to disperse their spores further (stay tuned for Part 2) and obtain more sunlight than competitors.

Recent major changes to the **embryophyte** tree of life have major implications for our understanding of early land plant evolution (Fig. 3). Historically, **bryophytes** were thought to be paraphyletic with respect to **polysporangiophytes**. In fact, the latest edition of the Raven textbook includes this arrangement. However, within the past few years (like basically yesterday), new molecular phylogenetic studies have converged on a phylogeny where **Bryophytes** are a monophyletic group, or clade. In this lab, we will consider the implications of these new results for understanding trait evolution along the **embryophyte** tree of life, and by exten-

sion, how we study the traits that lead to the success of land plants.

**Part 1 Goal:** Analyze the evolution of the sporic life cycle and alternation of generations across two phylogenetic hypotheses for Embryophytes.

## Exercise 1a: Evolution of Alternative Generations Under the Classic Tree

**Background:** Our first goal is to evaluate how the two different life history strategies involving alternation of generations - gametophyte and sporophyte dominant - evolved under the classic phylogenetic hypothesis - where bryophytes are paraphyletic. To do this we will use parsimony to reconstruct the evolution of the two alternative life histories on a phylogenetic tree that includes broad sampling of major embryophyte lineages.

**Exercise 1b Goals:** Reconstruct evolution of alternation of generations under the classic phylogenetic hypothesis involving paraphyly of bryophytes.

1. Open ITOI and upload the tree file - Lab4\_Exercise1a\_Tree1.new (Fig. 4). This file is a simple text document that contains a NEWICK formatted tree. Note: this lab includes several files with similar names, please be sure to carefully read and check the entire file name(s) to avoid the trouble that results from carelessly picking the wrong one (one of your instructors may have made this mistake while testing the lab).
2. Download and save the file that contains annotations for seven major clades of embryophytes - Lab4\_Exercise1a\_MajorClades.txt. Drag and drop this file into the ITOI window. You should see shaded boxes that indicate the following seven clades of embryophytes: mosses, hornworts, liverworts, lycophytes, ferns, gymnosperms, and angiosperms. You can view the names associated with each of these clades using the Datasets tab in the Controls window of ITOI (Fig. 4).
3. Download and save the file that contains data on whether the gametophyte is dominant or whether the sporophyte is



**Figure 3.** Two of the new studies - de Souza et al. 2018 and Harris et al. 2020 - that recover monophyly of Bryophytes.

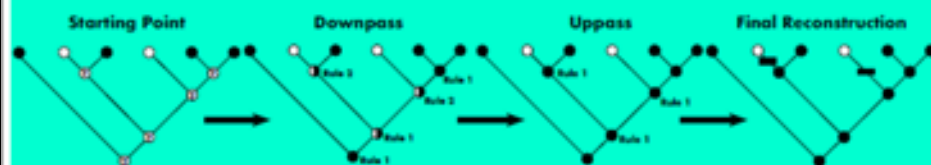
## Box 1: Reconstructing Evolution of Reversible Traits Using Fitch Parsimony

The goal of **parsimony** is to reconstruct the evolutionary scenario(s) with the fewest possible evolutionary changes. In the case of relatively small trees, where traits change infrequently, or where changes are irreversible, it is often possible to infer the most parsimonious solution "by eye" without the aid of any specific rules or algorithms (this is what you did last week with multicellularity). In many real world scenarios, however, we need to apply simple algorithms designed to efficiently identify the most parsimonious solution. The purpose of this box is to explain how to apply the **Fitch parsimony** algorithm.

The **Fitch parsimony** algorithm works by stepping through the tree one node at a time and making decisions about possible ancestral states that ensure the most parsimonious solution(s). The algorithm involves two steps. First, you complete an **downpass**, where possible ancestral states are reconstructed from the tips down to the root. During this downpass, the ancestral state at some internal nodes may be reconstructed as **ambiguous**, meaning that the node could be either a photosynthetic or non-photosynthetic. Second, you complete an **uppass** by working backwards from the root to the tips to resolve ambiguous nodes inferred during the downpass. During both the downpass and uppass, character states are reconstructed following two simple rules. Once you have completed the downpass and uppass, you can assign trait changes to branches in your tree.

**Rule 1:** If the descendants of an ancestral node share any common traits, assign those shared states to the ancestral node. Here are a few examples of how this rule is applied: (A) if your downpass starts with two descendants that are both photosynthetic, you would assign photosynthesis to the ancestor. (B) if one descendant is classified as ambiguous (i.e., either photosynthetic or non-photosynthetic) and the other as photosynthetic, you would assign photosynthesis to the ancestor. (C) if both descendants are classified as ambiguous, the ancestor would also be classified as ambiguous.

**Rule 2:** If the descendants of an ancestral node do not share traits in common traits, assign both of the possible states to the ancestor. For example, if one descendant has photosynthesis and the other does not, you would reconstruct the ancestor as ambiguous. For every node where you apply Rule 2, you will ultimately infer a trait change along one of the two descendant branches.



## Reflections teaching during Covid:

Class enrollment is virtually the same as compared to previous years

KU undergraduate biology program enrollment is up

Overall students have appreciated the flexibility in our class format and are learning

Rich and I are happy to share our lab material

## Outlook:

Specimen based research will continue

Fieldwork will resume next summer and beyond

Grant proposals focusing on collections based work are being written





## Taking the Pulse of Natural History Collections During COVID-19 Series: Where are we now?

### September 16: Collections' perspectives

*John Bates, The Field Museum*

*Emily Braker, University of Colorado Boulder*

*Brian Atkinson, The University of Kansas*

***Mare Nazaire, California Botanic Garden***



# California Botanic Garden's Collections and COVID: Creating the Silver Lining

Mare Nazaire, Herbarium Administrative Curator  
Chair, Collections Committee



## COVID-19: Challenges

- Budget constraints
- Reduced staffing
- No volunteers
- No fieldwork
- Delays: collecting permits
- Non-essential tasks on hold
- Staggered schedules
- Inside vs. outside work





Living Collection



# Seed Collection



# Library & Archives



# Herbarium

CANELLACEAE

DEGENERIACEAE

EUPOMATIACEAE

HERNANDIACEAE





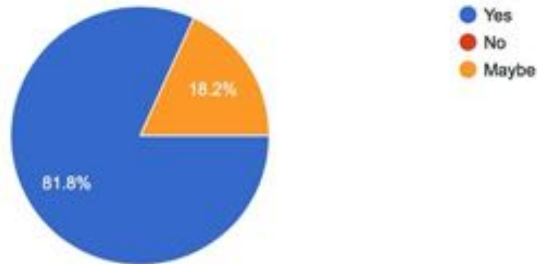




How do we connect students to collections when they cannot visit in person?

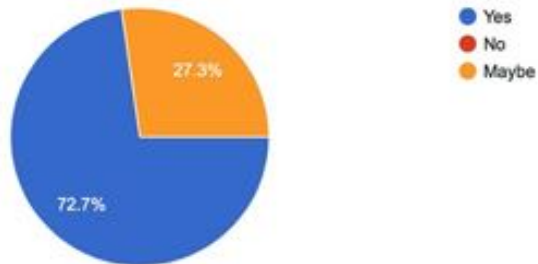
Would you be interested in supplementing your courses with a virtual (online) tour of the Garden's collections (living collection, library & archives, herbarium, and seed collection)?

11 responses



Would you be interested in supplementing your courses with online content about plant science and collections-based topics?

11 responses



### Surveyed the academic community:

- Previous visitation and tours
- Determine interest in virtual tours
- Determine interest in supplementing coursework





## **The Collections Connection**

Educational module:

*Plants in a changing world: California Diversity, Species Distributions, and Climate Change*

This work is supported by the Hispanic-Serving Institution's Education Grants Program, grant no. 2015-38422-24058/project accession no. 1007104, from the USDA National Institute of Food and Agriculture.



# CCH2

Featuring Data From the California Phenology  
TCN

Home Search Collections Map Search Checklists Image Search Browse Images Data Use Policy About CCH Welcome Here! My Profile Logout Settings

Home > Collections > Search Criteria > Specimen Records

Species List

Occurrence Records

Maps

Dataset: RSA-VascularPlants

Taxa: *Encolpopsis covillei* (*Encolpopsis argophylla* var. *grandiflora*, *Helianthella covillei*)

Search Criteria: has images; excluding cultivated/captive occurrences



1

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### RSA - California Botanic Garden Herbarium



*Encolpopsis covillei* (A. Nels.) Blake

RS40281889 S. C. Tschaez 4/1 1957-04-14

RSA-VascularPlants United States, California, Inyo County, Surprise Canyon (Balfour Quaternals), 36.11081°

N 117.19089, 762m

[Full Record Details](#)



1

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ASU Arizona State University  
Institutional Knowledge  
Integration Center  
www.asu.edu



### Explore projected long-term (30 year) Annual Average Maximum Temperature

ABSOLUTE VALUES

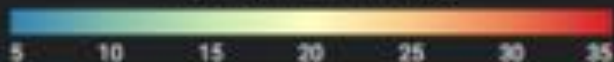
CHANGE FROM HISTORICAL BASELINE

Historical Baseline (1961–1990)

Mid-Century High Emissions (RCP 8.5)

#### Maps of Projected Change

Maximum Temperature (°C)



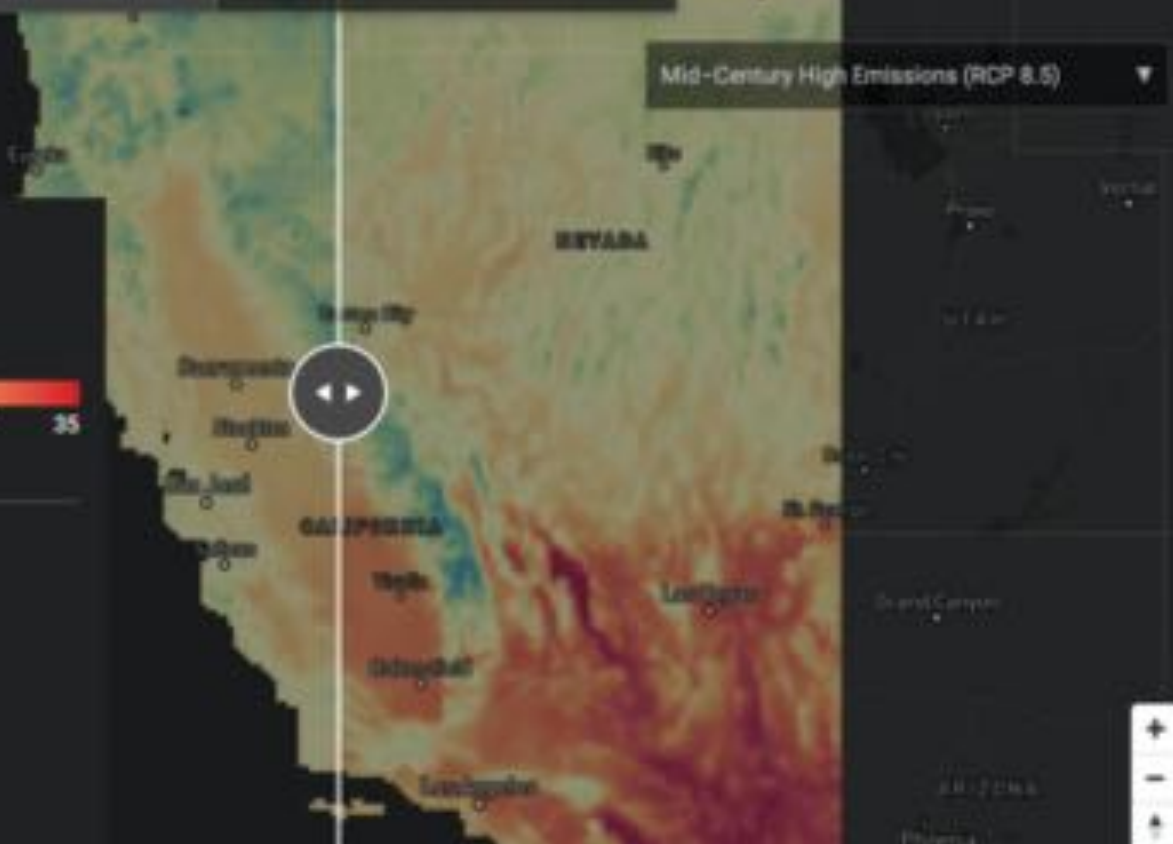
Change units:  Metric

Select boundary: None

Select map view:  SINGLE  SWIPE

Download: [IMAGES](#) [DATA](#)

[ABOUT THE TOOL](#)





For more info:  
<https://www.calbg.org>



## Break-out room discussion

What are the top three positive outcomes experienced by collections as we adapt to new COVID-aware workplace guidelines?

[bit.ly/idigbio-poll](https://bit.ly/idigbio-poll)

Break-outs (15 minutes) • Review (5 minutes) • Discussion



## Taking the Pulse of Natural History Collections During COVID-19 Series: Where are we now?

September 17: Moving Forward: Communities Response and Opportunities for the Future: Presentations and Panel Discussion/Q&A

*Speakers: Rob Gropp, Pam Soltis, Scott Miller, Roland Roberts*



Please help by providing feedback about this webinar series!

**Survey link:**

[https://ufl.qualtrics.com/jfe/form/SV\\_aVL7wEfLYUXcbFb](https://ufl.qualtrics.com/jfe/form/SV_aVL7wEfLYUXcbFb)